

In the Search of Genetic Polymorphism of Lactoferrin with PCR-RFLP in Imported Holsteins in a Large Tunisian Herd

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Abstract

The rate of subclinical mastitis inferred from somatic cell scores is high in Tunisian dairy cattle. Other than management, susceptibility of cows to infections may be of great impact on the mammary health. Lactoferrin seems to influence resistance of dairy cows to mastitis infections. The objective of this study was to search for genes coding for lactoferrin in imported Holstein cows managed in a large herd in the North of Tunisia. A total of 52 blood samples were collected. Genomic DNA was extracted and all individuals were genotyped for polymorphism of lactoferrin gene by PCR-RFLP analysis. Amplification of the lactoferrin gene fragment revealed an 1143 bp long product by electrophoresis following PCR. After restriction enzyme digestion with *HinfI*, two alleles were characterized by only one restriction fragment. All screened individuals were homozygous showing the same band pattern in all samples by the PCR-RFLP, implying the existence of only one genotype for the lactoferrin locus. This study may be generalized to cover the Tunisian Holstein population for possible identification of resistant animals to mastitis infection and that have polymorphic lactoferrin genes.

Keywords: Holstein cattle; lactoferrin gene; PCR-RFLP

Introduction

Rekik *et al.* (2008) reported that somatic cell scores (SCS) in the Tunisian Holstein population were high in the first three lactations most likely due to high mastitis infection rates. These infections resulted in reduced milk and protein yields and long calving to first service and calving to conception intervals. The SCS has high genetic correlation with mastitis (0.60-0.80), and is one of the useful indirect measures of mastitis at present (Zhang *et al.*, 2007).

Although sizeable genetic gains for health traits were made using conventional selection methods which are money and mainly time-consuming, there is a growing attention towards improving health traits using genomic selection in dairy populations worldwide (e.g., Schaeffer, 2006; de Roos *et al.*, 2010). In many countries which are part of Interbull, genomic selection is developing rapidly. Routine genomic evaluations produce genomically

enhanced breeding values (GEBV) of male calves which are only slightly less precise than conventional estimated breeding values (EBV) obtained after a formal progeny test. In such a context, several authors demonstrated that larger yearly genetic gains can be obtained by intensively using genomically evaluated young sires, without waiting for progeny tested results (Schaeffer, 2011).

In dairy cattle breeding special attention is given to candidate genes (alleles) and their association with production and health traits (Taylor *et al.*, 2006; Zhou *et al.*, 2006). Lactoferrin is an iron-binding glycoprotein found in most exocrine secretions including tears, saliva and milk, and there are numerous reports of its antibacterial activity *in vitro* and *in vivo* (Nibbering *et al.*, 2001; Wojdak *et al.*, 2006). The lactoferrin gene can be used as a marker of somatic cell concentration in milk and, in consequence, as a marker of susceptibility/resistance to mastitis in dairy cows.

Materials and Methods

Sample collection and DNA extraction

A total of 52 dairy cows were randomly selected from a private farm in the North of Tunisia. Blood samples were collected in vacutainer tubes containing EDTA (1 mg mL⁻¹). Genomic DNA was extracted using standard protocol (Easy-DNA™ Kit Invitrogen) and stored at -20°C until used in assay.

The concentration of DNA samples was estimated using UV-visible range spectrophotometer and diluted to 50 ng/μL before PCR amplification. All the DNA samples had 260/280 OD ratios in the range of 1.8 to 2, indicating high purity. DNA was also examined by loading samples on 0.8 % agarose gel and visualizing the band under UV light with a Gel Doc 1000 system (BioRad) after ethidium bromide staining.

The PCR was performed in a final volume of 50 μL containing 100 ng of template DNA, 50 pmole of each primer, 5 μL of 10X PCR buffer (20 mM Tris-HCl pH 8.4, 50 mM KCl), 1.5 mM MgCl₂, 0.2 mM of dNTPs, and 1 U of Taq DNA polymerase (Zhao *et al.*, 2008):

Forward: 5'-CACATTACAAGCAGGATCTTTTGCTG-3'
Reverse: 5'-CTGGCCAATGAGCCCTATATGTGT-3'

This solution was initially denatured at 94 °C for 5 min. followed by 39 cycles of denaturation (95 °C for 30 s), annealing (62 °C for 45 s), and elongation (72 °C for 1 min) and a final extension at 72 °C for 10 min. The PCR products were electrophoresed on 1.5% agarose gels in order to check the quality and specificity of DNA fragment amplification.

PCR-RFLP condition

For PCR-RFLP analysis, the 1143 bp PCR products were digested with *HinfI* (BioLabs). Restriction fragments were separated by electrophoresis in a 2% agarose gel and their sizes were estimated using the molecular markers. The results were taken into account when the sum of all the restriction fragments

for *HinfI* enzyme was in the range of 1143bp ± 100 (Fagundes and Dornelas, 2007). 20 μL of PCR products was digested for 4h at 37°C with 10 units of restriction enzyme. Digested products were separated by electrophoresis on a 1.5 % agarose gel and visualized with Ethidium bromide under UV light with a Gel Doc 1000 system (BioRad) after ethidium bromide staining.

Results and Discussion

Quality and quantity of extracted DNA from analyzed samples was tested by electrophoresis on agarose gel. Electrophoresis showed a height DNA quality. DNA quantity was estimated using the molecular markers quantity for each band. From all analyzed animals, DNA fragments containing the lactoferrin gene were amplified for nucleotide sequencing PCR using adequate primer pairs. As expected, the size of PCR product of lactoferrin gene was 1143 bp (Changhong *et al.*, 2009).

The results found show the existence of the sequence encoding lactoferrin in all sampled cows. Identification of different genotypes for lactoferrin requires an enzymatic digestion. In our case, digestion was carried out by restriction enzyme *HinfI*. This endonuclease cuts at the restriction site (5-G ↓ C-3 ANT, 3 - TNA C ↑ G -5) and subsequently allows the revelation of the restriction site if exists. Profile on agarose gel showed that all analyzed individuals were homozygous for the lactoferrin gene (Figure 1). All tested individuals showed a genotype A / A (a band size of 635 bp). The results found are similar to those found by Daly *et al.* (2006), Fagundes and Dornelas (2007), and O'Halloran *et al.* (2009).

After digestion, only the A/A genotype was obviously detected. The use of small samples from a private farm can explain results in this essay. According to Wojdak *et al.* (2006), individuals that are homozygous for the A allele presented the lowest rates of somatic cells. Similar observations were reported by Changhong *et al.* (2009) who confirmed that cows with the A/A genotypes were resistant to mastitis infection.

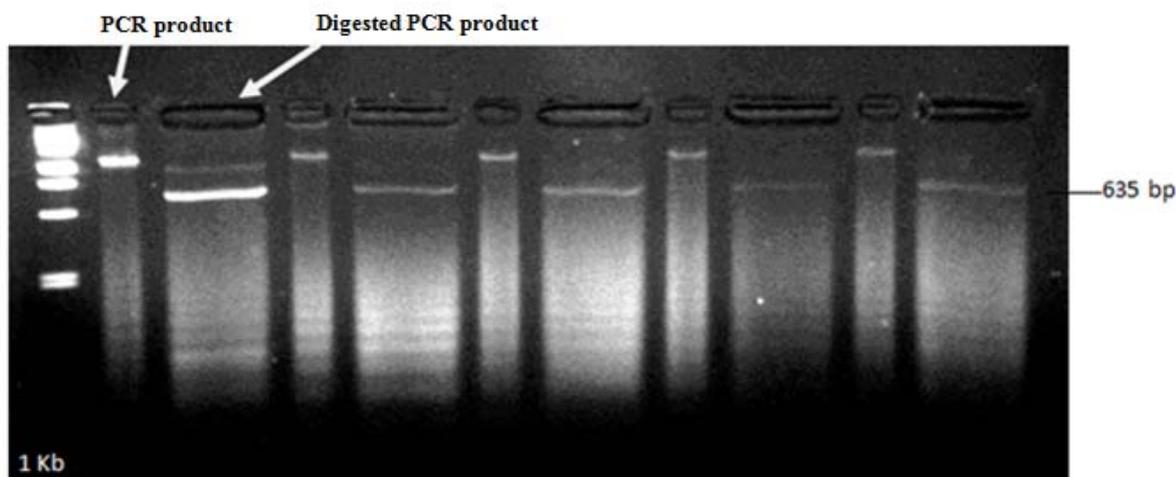


Figure 1. Detection of Lactoferrin variant by digested RFLP with *HinfI*. M: 1kb Molecular weight marker (invitrogen DNA Ladder).

Conclusion

Countries with currently no available genomic evaluations are often semen importers, the case of Tunisia. In the present essay, imported cows were sampled in a private Tunisian farm to search for polymorphism in the lactoferrin gene. The results of PCR-RFLP showed the same band pattern in all samples, implying one genotype for the lactoferrin locus. Only the A/A genotype was found in sampled animals. The A/A genotype for the lactoferrin was found to be associated with resistance to mastitis infections. This result can be used to design future studies to determine relations of lactoferrin alleles with resistance to mastitis in Tunisian Holsteins to collect data that may be used in genomic selection.

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