Quality and Value of Imputing Gene Tests for All Animals

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Abstract

Genomic selection is driven by genotyping arrays designed for uniform coverage of the genome because most quantitative trait loci (QTLs) underlying the heritability of the trait are unknown. Laboratories have improved the arrays since 2014 with custom content by adding selected QTLs discovered from whole-genome sequencing (WGS) and high-effect markers from higher-density arrays. Breed differences, missing data rates, and error rates were investigated for eight QTL gene tests currently imputed for all genotyped animals of 5 breeds plus crossbreds. Gene content for each breed was predicted for non-genotyped relatives using mixed model methods like those used in single-step genomic evaluations, allowing potential direct selection across all animals. For the 8 QTL studied, Mendel error rates were low except for polled in Jerseys and DGAT1 in most breeds. Allele effects for DGAT1 were smaller than two nearby flanking single nucleotide polymorphism (SNPs) because DGAT1 was genotype quality was poor on several arrays. For yield traits, 79K predictions including selected markers and QTLs had 1-2% higher reliability than 45K or 35K predictions excluding those SNPs.

Key words: Gene tests, imputation, marker selection, dairy cattle

Introduction

Genotyping laboratories began adding QTL gene tests in 2014 following the US Supreme Court decision that natural genetic variants should not be patented. Accuracy of imputing QTL genotypes for other animals can be affected by which arrays include the QTLs. Each year, new QTLs may be discovered and included. The SNP list used in US evaluations was updated frequently to include selected markers and QTLs from more breeds and higher density chips or from sequence (Al-Khudhair et al., 2021; Olson et al., 2012; VanRaden et al., 2009, 2017; Wiggans et al., 2016), with gains in reliability across traits expected to total about 3% (Table 1).

Some QTLs have effects larger than markers on traits we select or should select for. Goals of the project were to examine the most important QTLs currently used, summarize quality and breed differences of raw and imputed genotypes, estimate gene content for non-genotyped animals, and estimate gains in reliability of prediction from including or excluding the selected markers and gene tests.

Materials and Methods

Genotypes were examined from December 2022 official evaluations of the Council on Dairy Cattle Breeding (CDCB) for 5,669,157 Holstein, 663,366 Jersey, 65,172 Brown Swiss, 15,110 Ayrshire, and 7,620 Guernsey to summarize allele frequencies by breed (Table 2). Mendelian conflicts (Table 3) for eight important QTLs, and missing rates before and after imputation with DGAT1 as an example (Table 4). Gene content was estimated for all non-genotyped relatives by predicting their genotypes from relatives using Gengler (2007) method. To potentially include such QTLs in a selection index, non-genotyped candidates for selection also need estimates of their unknown QTLs.

For the QTLs studied (Table 5), some have economic merit not yet included in national selection indexes such as 1) polled mutations near 1:2578598 (chromosome: position on ARS-UCD1 map) that suppress horn growth, improve animal welfare, and reduce farm labor, 2) β-casein allele (a2) at 6:84451299 in a milk protein gene that may improve
digestibility, and 3) two \( \kappa \)-casein alleles near 6:84451299 that affect cheese yield. The three casein QTLs are in a 200kb gene duplication region. Other QTLs mainly affect traits already in selection such as 4) diacylglycerol O-acyltransferase 1 (DGAT1) at 14:611019 affecting fatty acid metabolism, percentages, and yields of fat and protein, 5) Bovine growth hormone receptor (BGHR) at 20:31888449 affecting protein percentage, 6) \( \beta \)-lactoglobulin (BLG) at 11:103259232 with large effects on yield especially in Brown Swiss, and 7) ATP binding cassette subfamily G member 2 (ABCG2) at 6:36599640 with the largest effect for milk, fat \%, protein \%, and net merit in Holsteins, but the favorable allele is now nearly fixed at 2.5\%, while fixed in other breeds (Table 2). Many other QTLs have recessive lethal effects and carrier status is reported, but those were not part of this study.

Genomic predictions using three SNP densities from 2019 yield trait data for 6,899 young Holstein bulls now proven allowed estimating the value of including selected markers and QTLs. The current 79K official list was compared to the 35K subset of only markers from the original 50K array and two 45K chips constructed by augmenting the 35K chip with independent sets of \( \frac{1}{4} \) of the high density (HD) SNPs, respectively.

**Results & Discussion**

A true QTL is expected to have a better genetic signal (effect size or genetic SD) compared to nearby markers on the chip and that was true for most QTLs. For Holsteins, the ABCG2 gene test had the best signal and the top ranked locus for milk, fat \%, protein \% and net merit. The BGHR gene test had the best signal and the second ranked locus for protein \%. But the DGAT1 gene test had a smaller effect than two nearby markers, and so attention was focused on DGAT1.

A locus from the 50K chip (ARS-BFGL-NGS-4939) on chromosome 14 at 609,870 bp had the largest genetic standard deviation (SD) genome-wide for the five Holstein yield traits: milk, fat, protein, fat \% and protein \%. That locus is 1,149 bp away from DGAT1, and another locus from the high-density chip (BovineHD1400000216) also had larger effects than DGAT1. Poor imputation quality was ruled out by comparing SNP regressions using only cows with direct calls for DGAT1 and the 50K SNP. Genotypes from nine of the 52 chips and 1,377,604 Holsteins had both loci, 46,051 (6\%) had discordant calls (gene test vs. marker), of which 6,830 had phenotypes. Six GeneSeek chips accounted for most of the data and had varying discordant rates (Table 6). The GeneSeek Genomic Profiler (GGP) 9K had the most genotyped animals (452,687), highest discordant rate (8.27\%), and 92\% (6281) of the phenotyped animals. GGP 9K regression effect sizes were greater and p-values smaller for the 50K SNP (Table 7). Genotype quality of GGP 9K was then assessed using SNP heritability (Gengler 2007) for 25,000 animals with discordant calls on that chip. The 50K SNP had heritability 0.98 and DGAT1 only 0.16, indicating poor genotype quality as the likely source. Discordant calls for DGAT1 on other chips also had low heritability although sample size was much smaller.

Because some valuable gene tests are sold by laboratories rather than delivered with array genotypes, freely imputed QTLs could benefit breeders and progress. Decreasing costs of whole genome sequence data will increase power of QTL discovery, and more QTL genotypes should increase imputation accuracy, prediction accuracy, and economic gain. Regressions averaged 1.07 and were nearly equal across the 3 densities. Reliabilities of yield traits for 79K averaged 1.2\% higher than 45K and 2.0\% higher than 35K, worth potentially > $10 million every year nationally. Eventually, more QTLs should be included to further improve predictions.
Conclusions

Gene tests were already imputed for all genotyped animals of all five breeds. Mendelian error rates were low for QTLs except for Polled in Jerseys and DGAT1 in most breeds. Imputed DGAT1 tests were statistically less significant for all yield traits compared to two nearby chip SNPs (one HD and one 50K), direct DGAT1 gene tests also had smaller effects than the best markers, and SNP heritability indicated that DGAT1 genotyping quality was the cause of later imputation errors, though the GGP 7K and linkage disequilibrium (LD) V4 had low discordance rates. Further investigation of problematic chips is warranted. Gene content was imputed for all non-genotyped animals by extracting QTLs from the imputed genotypes and using those as data to predict related animals. Accumulated gains in reliability for yield from adding selected markers and QTLs were 1-2%, a little less than previous studies indicated. Most gains were from larger reference populations.

Acknowledgments

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References


**Table 1.** History of US SNP list revisions and reported gains in reliability of Holstein predictions

<table>
<thead>
<tr>
<th>Year</th>
<th>Reference</th>
<th>Breeds</th>
<th>Added information</th>
<th>Markers (1000s)</th>
<th>HOL Reliability</th>
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</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Added</td>
<td>Total</td>
</tr>
<tr>
<td>&lt;2008</td>
<td>All</td>
<td>Parent average</td>
<td></td>
<td>0</td>
<td>27</td>
</tr>
<tr>
<td>2009</td>
<td>VanRaden</td>
<td>HO</td>
<td>Chip genotypes (50K)</td>
<td>38</td>
<td>38</td>
</tr>
<tr>
<td>2012</td>
<td>Olson</td>
<td>3</td>
<td>More breeds (JE, BS)</td>
<td>5</td>
<td>43</td>
</tr>
<tr>
<td>2013</td>
<td>Wiggans</td>
<td>HO</td>
<td>Add HD markers (GHD)</td>
<td>18</td>
<td>61</td>
</tr>
<tr>
<td>2016</td>
<td>Wiggans</td>
<td>HO</td>
<td>Add HD markers (GH2)</td>
<td>16</td>
<td>77</td>
</tr>
<tr>
<td>2019</td>
<td>VanRaden</td>
<td>HO</td>
<td>Add sequence SNPs</td>
<td>2</td>
<td>79</td>
</tr>
<tr>
<td>2020</td>
<td>Al-Khudhair</td>
<td>5</td>
<td>Add HD, other breeds</td>
<td>+5, -5</td>
<td>79</td>
</tr>
</tbody>
</table>

**Table 2.** Final allele frequencies for the eight QTLs including gene content for all animals of each breed

<table>
<thead>
<tr>
<th>Breed</th>
<th>Polled</th>
<th>ABCG2</th>
<th>β-casein</th>
<th>κ-casein1</th>
<th>κ-casein2</th>
<th>β-Lact</th>
<th>DGAT1</th>
<th>BGHR</th>
</tr>
</thead>
<tbody>
<tr>
<td>RDC</td>
<td>0.6</td>
<td>99.9</td>
<td>52.0</td>
<td>84.8</td>
<td>65.3</td>
<td>33.2</td>
<td>8.6</td>
<td>22.1</td>
</tr>
<tr>
<td>BSW</td>
<td>3.5</td>
<td>100.0</td>
<td>22.2</td>
<td>30.1</td>
<td>100.0</td>
<td>33.0</td>
<td>6.8</td>
<td>11.4</td>
</tr>
<tr>
<td>GUE</td>
<td>1.1</td>
<td>99.7</td>
<td>7.2</td>
<td>65.1</td>
<td>99.7</td>
<td>16.0</td>
<td>60.6</td>
<td>17.9</td>
</tr>
<tr>
<td>JER</td>
<td>2.2</td>
<td>99.9</td>
<td>27.6</td>
<td>9.2</td>
<td>99.4</td>
<td>54.2</td>
<td>52.1</td>
<td>26.1</td>
</tr>
<tr>
<td>HOL</td>
<td>1.0</td>
<td>97.4</td>
<td>39.1</td>
<td>72.5</td>
<td>89.8</td>
<td>51.6</td>
<td>30.1</td>
<td>19.7</td>
</tr>
</tbody>
</table>

**Table 3.** Mendelian error rates by breed for imputed genotypes of eight QTLs

<table>
<thead>
<tr>
<th>Breed</th>
<th>Polled</th>
<th>ABCG2</th>
<th>β-casein</th>
<th>κ-casein1</th>
<th>κ-casein2</th>
<th>β-Lact</th>
<th>DGAT1</th>
<th>BGHR</th>
</tr>
</thead>
<tbody>
<tr>
<td>RDC</td>
<td>0.01</td>
<td>0</td>
<td>0.17</td>
<td>0.00</td>
<td>0.01</td>
<td>0.05</td>
<td>0.80</td>
<td>0.11</td>
</tr>
<tr>
<td>BSW</td>
<td>0.18</td>
<td>0</td>
<td>0.10</td>
<td>0.12</td>
<td>0.00</td>
<td>0.12</td>
<td>0.51</td>
<td>0.03</td>
</tr>
<tr>
<td>GUE</td>
<td>0.00</td>
<td>0</td>
<td>0.00</td>
<td>0.04</td>
<td>0.00</td>
<td>0.14</td>
<td>0.00</td>
<td>0.07</td>
</tr>
<tr>
<td>JER</td>
<td>0.50</td>
<td>0</td>
<td>0.17</td>
<td>0.13</td>
<td>0.00</td>
<td>0.03</td>
<td>0.09</td>
<td>0.08</td>
</tr>
<tr>
<td>HOL</td>
<td>0.05</td>
<td>0</td>
<td>0.08</td>
<td>0.01</td>
<td>&lt;0.01</td>
<td>0.02</td>
<td>0.67</td>
<td>0.10</td>
</tr>
</tbody>
</table>

**Table 4.** DGAT1 imputed allele and genotype frequencies and genotypes missing in input

<table>
<thead>
<tr>
<th>Breed</th>
<th>Tests (N)</th>
<th>Allele</th>
<th>Imputed genotype codes</th>
<th>Genotypes</th>
<th>Frequency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>RDC</td>
<td>15,110</td>
<td>8.6</td>
<td>88.11</td>
<td>8.84</td>
<td>0.07</td>
</tr>
<tr>
<td>BSW</td>
<td>65,172</td>
<td>6.8</td>
<td>78.14</td>
<td>9.27</td>
<td>0.45</td>
</tr>
<tr>
<td>GUE</td>
<td>7,620</td>
<td>60.6</td>
<td>14.00</td>
<td>43.24</td>
<td>33.23</td>
</tr>
<tr>
<td>JER</td>
<td>663,366</td>
<td>52.1</td>
<td>21.34</td>
<td>49.43</td>
<td>27.63</td>
</tr>
<tr>
<td>HOL</td>
<td>5,669,157</td>
<td>30.1</td>
<td>46.10</td>
<td>42.70</td>
<td>9.60</td>
</tr>
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</table>
### Table 5. Locations and effects of eight QTLs examined

<table>
<thead>
<tr>
<th>Gene test</th>
<th>Chr:Location</th>
<th>Gene function</th>
<th>Effects in cows or in humans</th>
</tr>
</thead>
<tbody>
<tr>
<td>Polled</td>
<td>1:2578598</td>
<td>Grow horns</td>
<td>Animal welfare, farm labor</td>
</tr>
<tr>
<td>ABCG2</td>
<td>6:36599640</td>
<td>Membrane transport</td>
<td>Yield and NM$ (biggest effect)</td>
</tr>
<tr>
<td>(\beta)-casein (a2)</td>
<td>6:84451299</td>
<td>Milk protein</td>
<td>More digestible? (JE protein%)</td>
</tr>
<tr>
<td>K-casein (1)</td>
<td>6:85656772</td>
<td>Milk protein</td>
<td>Increased cheese yield</td>
</tr>
<tr>
<td>K-casein (2)</td>
<td>6:85656792</td>
<td>Milk protein</td>
<td>Increased cheese yield</td>
</tr>
<tr>
<td>(\beta)-Lactoglobulin</td>
<td>11:103259232</td>
<td>Milk fat</td>
<td>Human allergies (BS yield &amp; %)</td>
</tr>
<tr>
<td>DGAT1</td>
<td>14:611019</td>
<td>Fat and protein %</td>
<td>Fatty acid metabolism, obesity</td>
</tr>
<tr>
<td>BGHR</td>
<td>20:31888449</td>
<td>Growth hormone</td>
<td>Protein% (2nd biggest effect)</td>
</tr>
</tbody>
</table>

### Table 6. Descriptive statistics for six GeneSeek chips tested for DGAT1 calling

<table>
<thead>
<tr>
<th>Chip info</th>
<th>Animal info</th>
</tr>
</thead>
<tbody>
<tr>
<td>Name</td>
<td>Markers</td>
</tr>
<tr>
<td>GGP 7K</td>
<td>7083</td>
</tr>
<tr>
<td>GGP 9K</td>
<td>8984</td>
</tr>
<tr>
<td>GGP LD V4</td>
<td>30113</td>
</tr>
<tr>
<td>GGP 65K</td>
<td>65320</td>
</tr>
<tr>
<td>GGP 100K</td>
<td>94121</td>
</tr>
<tr>
<td>GGP 150K</td>
<td>139914</td>
</tr>
</tbody>
</table>

### Table 7. Regression results for GGP 9K chip for DGAT1 vs. nearby 50K SNP using 6,281 genotyped animals

<table>
<thead>
<tr>
<th>Marker</th>
<th>P-value</th>
<th>Abs (marker effect)</th>
<th>50K</th>
<th>DGAT1</th>
<th>50K</th>
<th>DGAT1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Milk</td>
<td>8.9E-45</td>
<td>2.6E-02</td>
<td>70.932</td>
<td>11.887</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fat</td>
<td>1.4E-19</td>
<td>3.1E-02</td>
<td>2.062</td>
<td>0.546</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protein</td>
<td>4.9E-13</td>
<td>9.8E-01</td>
<td>0.967</td>
<td>0.004</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fat %</td>
<td>2.2E-04</td>
<td>3.8E-04</td>
<td>0.016</td>
<td>0.003</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protein %</td>
<td>4.1E-02</td>
<td>1.2E-04</td>
<td>0.003</td>
<td>0.001</td>
<td></td>
<td></td>
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</tbody>
</table>